



Granzyme B ELISpot Assay Kit (Human)

Instructions for use

Catalogue Numbers:

| | Pre-coated |
|------------|------------|
| 1x96 tests | OKDB00023 |
| 5x96 tests | OKDB00024 |

For research use only

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Human Granzyme B ELISpot Kit – Pre-coated

1. Intended use

Aviva **ELISpot** is a highly specific immunoassay for the analysis of cytokine and other soluble molecule production and secretion from T-cells at a single cell level in conditions closely comparable to the *in-vivo* environment with minimal cell manipulation. This technique is designed to determine the frequency of cytokine producing cells under a given stimulation and the comparison of such frequency against a specific treatment or pathological state. The ELISpot assay constitutes an ideal tool in the investigation of Th1 / Th2 responses, vaccine development, viral infection monitoring and treatment, cancerology, infectious disease, autoimmune diseases and transplantation.

Utilising sandwich immuno-enzyme technology, Aviva ELISpot assays can detect both secreted cytokines and single cells that simultaneously produce multiple cytokines. Cell secreted cytokines or soluble molecules are captured by coated antibodies avoiding diffusion in supernatant, protease degradation or binding on soluble membrane receptors. After cell removal, the captured cytokines are revealed by tracer antibodies and appropriate conjugates.

This kit has been configured for research use only and is not to be used in diagnostic procedures.

2. Introduction

2.1. Summary

Granzyme B belongs to the granzymes family, which are exogenous serine proteinases. Granzymes are produced by cytotoxic lymphocytes (CTLs) and NK cells

Activated cytotoxic T lymphocytes and NK cells are important effectors in immunologic defenses against tumor cell proliferation, viral infection and transplant tissue. They induce apoptosis via multiple mechanism thanks to granules exocytosis. Following adhesion of the CTL on the target cell (via CTL-receptor and antigen-presenting MHC molecules), CTLs secrete cytoplasmic granules containing granules and enzymes including granzymes and perforine (pore-forming protein). Granules are then released in the intercellular space, perforine forms a transmembrane pore .Through these pores, granzymes enter the cytosol of the target cell.

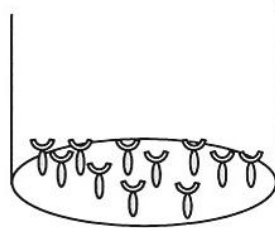
Granzyme B is the most powerful pro-apoptotic member of granzyme family. Granzyme B activates the intracellular caspases inducing finally the death of the cell.


Granzymes are diagnostics markers for many disorders (viral infection, lymphoma's and carcinoma's, auto-immune disease and rejection of transplant).

2.2. Principle of the method

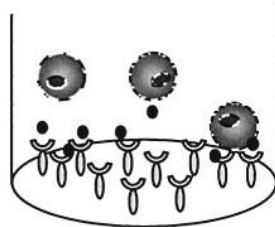
A capture antibody highly specific for the analyte of interest is coated to the wells of a PVDF bottomed 96 well microtitre plate either during kit manufacture or in the laboratory. The plate is then blocked to minimise any non-antibody dependent unspecific binding and washed. Cell suspension and stimulant are added and the plate incubated allowing the specific antibodies to bind any analytes produced. Cells are then removed by washing prior to the addition of Biotinylated detection antibodies which bind to the previously captured analyte. Enzyme conjugated streptavidin is then added binding to the detection antibodies. Following incubation and washing substrate is then applied to the wells resulting in coloured spots which can be quantified using appropriate analysis software or manually using a microscope.

1. 96-PVDF bottomed-well plates are first treated with 35% ethanol and then coated with capture antibody





 Capture antibody


2. Incubation of cells in the coated microwell



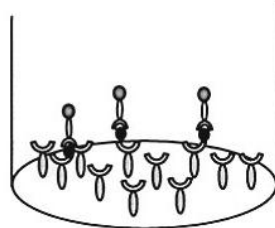
 Antigen / Mitogen

 Biotinylated detection antibody

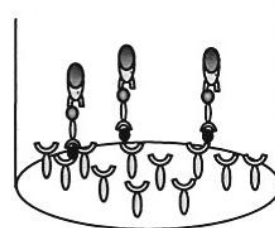
 Streptavidin - alkaline phosphatase conjugated

 Substrate product

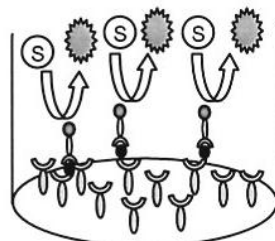
3. Cell removal by washing. Incubation with biotinylated antibody



4. Incubation with streptavidin – alkaline phosphatase conjugated



5. Addition of substrate BCIP/NBT and monitoring of spot formation.



3. Reagents provided (Contents shown for 5x96 test format)

- Pre-coated 96 well PVDF bottomed plates (5)
- Biotinylated detection antibody (lyophilised, resuspend in 0.55ml)
- Streptavidin-Alkaline Phosphatase conjugate (50µl)
- Bovine Serum Albumin (BSA)
- Ready to use BCIP/NBT substrate buffer (50ml)

Please note for 1x96 demo kits, Biotinylated detection antibody is provided in liquid form.

4. Materials/Reagents required but not provided

- Miscellaneous laboratory plastic and/or glass, if possible sterile
- Cell culture reagents (e.g. RPMI-1640, L-glutamine, FCS)
- Cell stimulation reagents (e.g. PHA, PMA, Ionomycin)
- CO₂ incubator
- Tween 20
- Phosphate Buffered Saline (PBS)

5. Storage Instructions

Store kit reagents between 2 and 8°C. Immediately after use remaining reagents should be returned to cold storage (2 to 8°C). Expiry of the kit and reagents is stated on box front labels. The expiry of the kit components can only be guaranteed if the components are stored properly, and if in the case of repeated use of one component, the reagent is not contaminated by the first handling.

6. Safety & Precautions for use

- For **research use only** not to be used as a diagnostic test
- Handling of reagents, serum or plasma specimens should be in accordance with local safety procedures , e.g.CDC/NIH Health manual : " Biosafety in Microbiological and Biomedical Laboratories" 1984
- Do not eat, drink, smoke or apply cosmetics where kit reagents are used
- Do not pipette by mouth
- When not in use, kit components should be stored refrigerated or frozen as indicated on vials or bottles labels
- All reagents should be warmed to room temperature before use.
- Cover or cap all reagents when not in use
- Do not mix or interchange reagents between different lots
- Do not use reagents beyond the expiration date of the kit
- Use a clean disposable plastic pipette tip for each reagent, standard, or specimen addition in order to avoid cross contamination
- Use a clean plastic container to prepare the washing solution
- Thoroughly mix the reagents and samples before use by agitation or swirling
- All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells
- When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells
- **BCIP/NBT buffer** is potentially carcinogenic and should be disposed of appropriately, caution should be taken when handling this reagent, always wear gloves
- Follow incubation times described in the assay procedure

7. Reagent Preparation

7.1. 1X Phosphate Buffered Saline (PBS)

For 1 litre of 10X PBS weigh-out: 80g NaCl
 2g KH₂PO₄
 14.4g Na₂HPO₄ · 2H₂O.

Add distilled water to 1 litre. Adjust the pH of the solution to 7.4 +/- 0.1 were required.

Dilute the solution to 1X before use.

7.2. 1% BSA PBS Solution (Dilution Buffer)

For one plate dissolve 0.2 g of BSA in 20 ml of 1X PBS.

7.3. 0.05% PBS-T Solution (Wash Buffer)

For one plate dissolve 50µl of Tween 20 in 100 ml of 1X PBS.

7.4. Detection Antibody

Reconstitute the lyophilised antibody with 0.55mL of distilled water. Gently mix the solution and wait until all the lyophilised material is back into solution.

Please note for 1x96 demo kits, Biotinylated detection antibody is provided in liquid form.

If not used within a short period of time, reconstituted Detection Antibody should be aliquoted and stored at -20°C. In these conditions the reagent is stable for at least one year. For optimal performance prepare the reconstituted antibody dilution immediately prior to use.

Dilute 100µl of antibody into 10ml Dilution Buffer and mix well.

For optimal performance, filter the solution using a disposable syringe and a 0.2µm filter disc.

7.5. Streptavidin – AP conjugate

For optimal performance prepare the Streptavidin-AP dilution immediately prior to use

For 1 plate dilute 10µl of Streptavidin-AP conjugate into 10 mL Dilution Buffer and mix well.

Do not keep this solution for further experiments.

For optimal performance, filter the Streptavidin solution using a disposable syringe and a 0.2µm filter disc.

7.6. BCIP / NBT

This reagent is ready-to-use.

In case of precipitation, filter the solution using a disposable syringe and a 0.2µm filter disc to avoid the presence of artifactual spots.

8. Sample and Control Preparation

8.1. Cell Stimulation

Cells can either be stimulated directly in the antibody coated wells (Direct) or, first stimulated in 24 well plates or flask, harvested, and then plated into the coated wells (Indirect).

The method used is dependent on 1) the type of cell assayed 2) the expected cell frequency. When a low number of cytokine producing cells are expected it is also advised to test them with the direct method, however, when this number is particularly high it is better to use the indirect ELISpot method.

All the method steps following stimulation of the cells are the same whatever the method (direct/indirect) chosen.

8.2. Positive Assay Control, Granzyme B production

We recommend using the following polyclonal activation as a positive control in your assay.

Dilute PBMC in culture media (e.g. RPMI 1640 supplemented with 2mM L-glutamine and 10% heat inactivated fetal calf serum) containing 1ng/ml PMA and 500ng/ml ionomycin (Sigma, Saint Louis, MO). Distribute 1×10^5 to 2.5×10^5 cells per 100 μ l in required wells of an antibody coated 96-well PVDF plates and incubate for 15-20 hours in an incubator.

For other stimulators incubation times may vary, depending on the frequency of cytokine producing cells, and should be optimised in each situation.

8.3. Negative Assay Control

Dilute PBMC in culture media to give an appropriate cell number (same number of unstimulated cells as stimulated sample cells) per 100 μ l with no stimulation.

8.4. Sample

Dilute PBMC in culture medium and stimulator of interest (i.e. Sample, Vaccine, Peptide pool or infected cells) to give an appropriate cell number per 100 μ l.

Optimal assay performances are observed between 1×10^5 and 2.5×10^5 cells per 100 μ l.

Stimulators and incubation times can be varied depending on the frequency of cytokine producing cells and therefore should be optimised by the testing laboratory.

9. Method

Prepare all reagents as shown in section 7 and 8.

Note: For optimal performance prepare the Streptavidin-AP dilution immediately prior to use

| Assay Step | | Details |
|--|-------------|--|
| 1. | Addition | Add 100µl of 1X PBS to every well |
| 2. | Incubation | Incubate plate at room temperature (RT) for 10 min |
| 3. | Wash | Empty the wells by flicking the plate over a sink & gently tapping on absorbent paper. |
| 4. | Addition | Add 100µl of sample, positive and negative controls cell suspension to appropriate wells providing the required concentration of cells and stimulant (cells may have been previously stimulated see section 8.) |
| 5. | Incubation | Cover the plate and incubate at 37°C in a CO ₂ incubator for an appropriate length of time (10-15 hours) Note: do not agitate or move the plate during this incubation |
| 6. | Addition | Empty the wells and remove excess solution then add 100µl of PBS-T to every well |
| 7. | Incubation | Incubate the plate at 4°C for 10 min |
| 8. | Wash | Empty the wells as previous and wash the plate 3x with 100µl of PBS-T |
| 9. | Addition | Add 100µl of diluted detection antibody to every well |
| 10. | Incubation | Cover the plate and incubate at 37°C for 1 hour 30 min |
| 11. | wash | Empty the wells as previous and wash the plate 3x with 100µl of PBS-T |
| 12. | Addition | Add 100µl of diluted Streptavidin-AP conjugate to every well |
| 13. | Incubation | Cover the plate and incubate at room temperature for 1 hour |
| 14. | Wash | Empty the wells and wash the plate 3x with 100µl of PBS-T |
| 15. | Wash | Peel of the plate bottom and wash both sides of the membrane 3x under running distilled water, once washing complete remove any excess solution by repeated tapping on absorbent paper. |
| 16. | Addition | Add 100µl of ready-to-use BCIP/NBT buffer to every well |
| 17. | Development | Incubate the plate for 5-15 min monitoring spot formation visually throughout the incubation period to assess sufficient colour development |
| 18. | Wash | Empty the wells and rinse both sides of the membrane 3x under running distilled water. Completely remove any excess solution by gentle repeated tapping on absorbent paper |
| <p>Read Spots: allow the wells to dry and then read results. The frequency of the resulting coloured spots corresponding to the cytokine producing cells can be determined using an appropriate ELISpot reader and analysis software or manually using a microscope.</p> <p><i>Note: spots may become sharper after overnight incubation at 4°C</i></p> | | |

Plate should be stored at RT away from direct light, but please note colour may fade over prolonged periods so read results within 24 hours.

10. Performance Characteristics

10.1. Reproducibility and Linearity

Intra-assay reproducibility and linearity were evaluated by measuring the spot development following the stimulation (PMA / Ionomycin) of 6 different PBMC cell concentrations, 12 repetitions in 1 batch. The data shows the mean spot number, range and CV for the six cell concentrations.

| Cells / well | n | Mean number of spots per well | Min | Max | CV% |
|--------------|----|-------------------------------|-----|-----|-------|
| 100000 | 12 | 835 | 796 | 864 | 2.6% |
| 50000 | 12 | 668 | 638 | 708 | 2.9% |
| 25000 | 12 | 474 | 425 | 499 | 4.2% |
| 12500 | 12 | 304 | 277 | 334 | 5.6% |
| 6250 | 12 | 155 | 106 | 193 | 16.8% |
| 3125 | 12 | 91 | 63 | 101 | 11.1% |

11. Bibliography

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2. Otake S, Kam CM, Narasimhan L, Poe M, Blake JT, Krahenbuhl O, Tschopp J, Powers JC, (1991). Human and murine cytotoxic T lymphocyte serine proteases: subsite mapping with peptide thioester substrates and inhibition of enzyme activity and cytolysis by isocoumarins. *Biochemistry.* 1991 Feb 26;30(8):2217-27.
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5. Spaeny-Dekking EH, Hanna WL, Wolbink AM, Wever PC, Kummer AJ, Swaak AJ, Middeldorp JM, Huisman HG, Froelich CJ, Hack CE, (1998). Extracellular granzymes A and B in humans: detection of native species during CTL responses in vitro and in vivo. *J Immunol.* 1998 Apr 1;160(7):3610-6.

12. Aviva Granzyme B ELISpot references

- Biagi E. et al., *Blood*, 2005; 105(6): 2436 – 2442.
Godard B. et al., *Hum. Immunol.*, 2004; 65(11): 1307-18.
Lindner, S. et al., *Cancer Res.*, 2013; 73(8): 2468-2479.

13. Technical Resources

Technical Support:

For optimal service please be prepared to supply the lot number of the kit used.

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