

**LEP ELISA Kit (Monkey)**  
**OKCA02323**  
**Instructions for Use**

For the quantitative measurement of Monkey LEP in serum, plasma and tissue homogenates.

Lot to lot variation can occur. Refer to the manual provided with the kit.

This product is intended for research use only.

## Table of Contents

1. Background.....	2
2. Assay Summary .....	3
3. Storage and Stability .....	3
4. Kit Components.....	3
5. Precautions .....	4
6. Required Materials Not Supplied .....	4
7. Technical Application Tips.....	4
8. Reagent Preparation .....	5
9. Sample Preparation.....	7
10. Assay Procedure.....	8
11. Calculation of Results.....	9
12. Typical Expected Data .....	9
13. Technical Resources.....	10

## 1. Background

### Principle

Aviva Systems Biology LEP ELISA Kit (Monkey) (OKCA02323) is based on a competitive enzyme immuno assay technique. The microtiter well-plate in this kit has been pre-coated with LEP. Sample or standards are added to the wells along with a fixed quantity of HRP-Conjugated LEP Detector Antibody and incubated. The XYLT1 found in the sample or standards competes with immobilized LEP for binding with the HRP-Conjugated LEP Detector Antibody. Wells are washed and an enzymatic reaction is then produced through the addition of TMB substrate which is catalyzed by the immobilized HRP to generate a blue color product. Addition of acidic stop solution changes the coloration to yellow. The density of yellow coloration is measured by reading the absorbance at 450 nm which is quantitatively proportional to the amount of biotinylated XYLT1 captured in the well and inversely proportional to the amount of XYLT1 which was contained in the sample or standard.

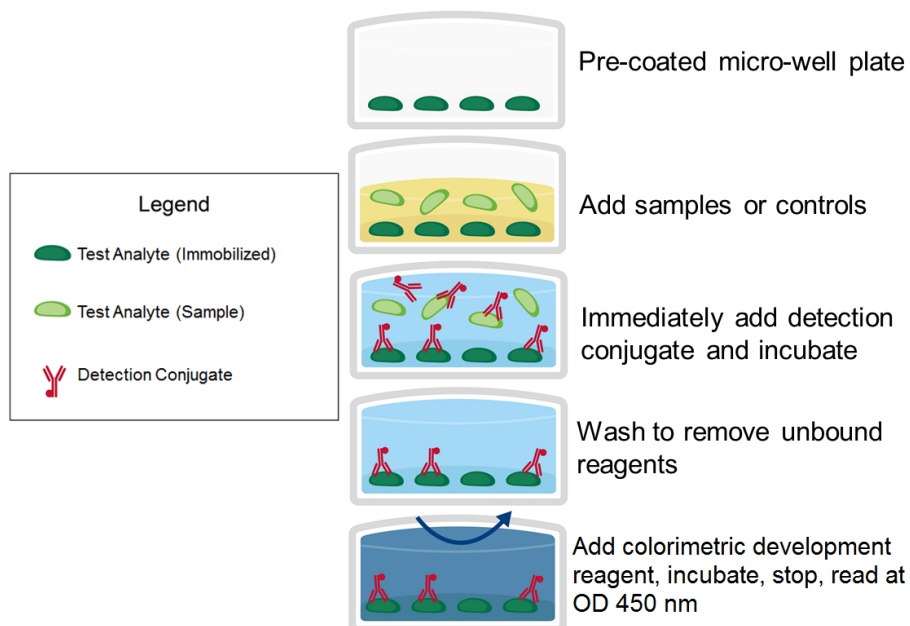
### Target Background

Key player in the regulation of energy balance and body weight control. Once released into the circulation, has central and peripheral effects by binding LEPR, found in many tissues, which results in the activation of several major signaling pathways. In the hypothalamus, acts as an appetite-regulating factor that induces a decrease in food intake and an increase in energy consumption by inducing anorexigenic factors and suppressing orexigenic neuropeptides, also regulates bone mass and secretion of hypothalamo-pituitary-adrenal hormones. In the periphery, increases basal metabolism, influences reproductive function, regulates pancreatic beta-cell function and insulin secretion, is pro-angiogenic for endothelial cell and affects innate and adaptive immunity. In the arcuate nucleus of the hypothalamus, activates by depolarization POMC neurons inducing FOS and SOCS3 expression to release anorexigenic peptides and inhibits by hyperpolarization NPY neurons inducing SOCS3 with a consequent reduction on release of orexigenic peptides. In addition to its known satiety inducing effect, has a modulatory role in nutrient absorption. In innate immunity, modulates the activity and function of neutrophils by increasing chemotaxis and the secretion of oxygen radicals. Increases phagocytosis by macrophages and enhances secretion of pro-inflammatory mediators. Increases cytotoxic ability of NK cells. Plays a pro-inflammatory role, in synergy with IL1B, by inducing NOS2 which promotes the production of IL6, IL8 and Prostaglandin E2, through a signaling pathway that involves JAK2, PI3K, MAP2K1/MEK1 and MAPK14/p38. In adaptive immunity, promotes the switch of memory T-cells towards T helper-1 cell immune responses. Increases CD4+CD25- T-cell proliferation and reduces autophagy during TCR (T-cell receptor) stimulation, through MTOR signaling pathway activation and BCL2 up-regulation.

### General Specifications

General Specifications	
Range	3.125 ng/mL- 50 ng/mL
LOD	3.125 ng/mL (Derived by linear regression of OD <sub>450</sub> of the Mean Blank + 2xSD)
Specificity	<p>Monkey Leptin</p> <p><u>UniProt</u>: Q28504</p> <p><u>GeneID</u>: 698728</p> <p><u>Target Alias</u>: FLJ94114, OB, OBS, leptin (murine obesity homolog) leptin (obesity homolog, mouse) obese, mouse, homolog of obesity factor</p>
Cross-Reactivity	No detectable cross-reactivity with other relevant proteins

## 2. Assay Summary



## 3. Storage and Stability

- Upon receipt store kit at 4°C for 1 month. Do not use beyond expiration date.

## 4. Kit Components

- The following reagents are the provided contents of the kit.

Description	Quantity	Storage Conditions
Anti-LEP Microplate	96 Wells (12 x 8 Well strips)	4°C for 1 Month  Do not use past expiration date.
LEP Lyophilized Standard	5 x 1 mL	
100X HRP-Conjugated LEP Detector Antibody	1 x 6 mL	
HRP-Conjugate Diluent	1 x 6 mL	
20X Wash Buffer	1 x 15 mL	
Substrate A	1 x 7 mL	
Substrate B	1 x 7 mL	
Stop Solution	1 x 7 mL	

## 5. Precautions

- Read instructions fully prior to beginning use of the assay kit.
- Any deviations or modifications from the described method or use of other reagents could result in a reduction of performance.
- Reduce exposure to potentially harmful substances by wearing personal protective lab equipment including lab coats, gloves and glasses.
- For information on hazardous substances included in the kit please refer to the Material Safety Data Sheet (MSDS).
- Kit cannot be used beyond the expiration date on the label.

## 6. Required Materials Not Supplied

- Microplate reader capable of reading absorbance at 450 nm.
- Automated plate washer (optional).
- Pipettes capable of precisely dispensing 0.5  $\mu$ L through 1 mL volumes of aqueous solutions.
- Pipettes or volumetric glassware capable of precisely measuring 1 mL through 100 mL of aqueous solutions.
- New, clean tubes and/or micro-centrifuge tubes for the preparation of standards or samples.
- Absorbent paper or paper toweling.
- Distilled or deionized ultrapure water.
- 37°C Incubator (optional)

## 7. Technical Application Tips

- Do not mix or substitute components from other kits.
- To ensure the validity of experimental operation, it is recommended that pilot experiments using standards and a small selection of sample dilutions to ensure optimal dilution range for quantitation.
- Samples exhibiting OD measurements higher than the highest standard should be diluted further in the appropriate sample dilution buffers.
- Prior to using the kit, briefly spin component tubes to collect all reagents at the bottom.
- Replicate wells are recommended for standards and samples.
- Cover microplate while incubating to prevent evaporation.
- Do not allow the microplate wells dry at any point during the assay procedure.
- Do not reuse tips or tube to prevent cross contamination.
- Avoid causing bubbles or foaming when pipetting, mixing or reconstituting.
- Completely remove of all liquids when washing to prevent cross contamination.
- Prepare reagents immediately prior to use and do not store, with the exception of the top standard.
- Equilibrate all materials to ambient room temperature prior to use (standards exception).
- For optimal results in inter- intra- assay consistency, equilibrate all materials to 37°C prior to performing assay (standards exception) and perform all incubations at 37°C.
- Pipetting less than 1  $\mu$ L is not recommended for optimal assay accuracy.
- Once the procedure has been started, all steps should be completed without interruption. Ensure that all reagents, materials and devices are ready at the appropriate time.
- Incubation times will affect results. All wells should be handled in the same sequential order and time intervals for optimal results.
- Samples containing precipitates or fibrin strands or which are hemolytic or lipemic might cause inaccurate results due to interfering factors.
- TMB Substrate is easily contaminated and should be colorless or light blue until added to plate. Handle carefully and protect from light.

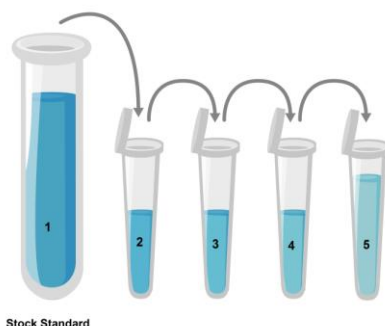
## 8. Reagent Preparation

- Equilibrate all materials to room temperature prior to use and use prepare immediately prior to use.

### 8.1 Monkey LEP Assay Standards

- 8.1.1 Prepare the LEP standards no greater than 2 hours prior to performing experiment. Standards should be held on ice until use in the experiment.
- 8.1.2 Reconstitute one vial of the provided 50 ng **Lyophilized Standard** for each experiment. Prepare a stock **50 ng/mL Standard** by reconstituting one tube of 50 ng **Lyophilized Standard** as follows:
- 8.1.2.1 Gently spin or tap the vial at 6,000 – 10,000 rpm for 30 seconds to collect all material at the bottom.
- 8.1.2.2 Add 1 mL of **Standard Diluent** to the vial.
- 8.1.2.3 Seal the vial then mix gently and thoroughly.
- 8.1.2.4 Leave the vial at ambient temperature for 15 minutes.
- 8.1.3 Prepare a set of serially diluted standards as follows:
- 8.1.3.1 Label tubes with numbers 2 – 6.
- 8.1.3.2 Use the undiluted reconstituted **50 ng/mL Standard** from step 8.1.2 as the high standard point (Tube #1).
- 8.1.3.3 Add 300  $\mu$ L of **Standard Diluent** to Tube #'s 2 – 6.
- 8.1.3.4 Prepare **Standard #2** by adding 300  $\mu$ L of **50 ng/mL Standard** (Tube #1) to Tube #2. Mix gently and thoroughly.
- 8.1.3.5 Prepare **Standard #3** by adding 300  $\mu$ L of **Standard #2** from Tube #2 to Tube #3. Mix gently and thoroughly.
- 8.1.3.6 Prepare further serial dilutions through Tube #6. Reference the table below as a guide for serial dilution scheme.
- 8.1.3.7 Tube #6 is a blank standard (only **Standard Diluent**), which should be included with every experiment.

Standard Number (Tube)	Standard To Dilute	Volume Standard to Dilute ( $\mu$ L)	Volume Standard Diluent Buffer ( $\mu$ L)	Total Volume ( $\mu$ L)	Final Concentration
1	50 ng/mL Reconstituted Standard	NA	NA	NA	50 ng/mL
2	50 ng/mL	300	300	600	25 ng/mL
3	25 ng/mL	300	300	600	12.5 ng/mL
4	12.5 ng/mL	300	300	600	6.25 ng/mL
5	6.25 ng/mL	300	300	600	3.125 ng/mL
6	NA	0	300	300	0.0 (Blank)



## 8.2 **1X HRP Conjugated LEP Antibody**

- 8.2.1 Prepare the **1X HRP-Conjugated LEP Detector Antibody** immediately prior to use by reconstituting and diluting the **100X HRP-Conjugated LEP Detector Antibody** 1:100 with **HRP-Conjugate Diluent** as follows.
- 8.2.2 Equilibrate materials to room temperature.
- 8.2.3 For each well strip to be used in the experiment (8-wells) prepare 500  $\mu\text{L}$  of **1X HRP-Conjugated LEP Detector Antibody** by adding 5  $\mu\text{L}$  of **100X HRP-Conjugated LEP Detector Antibody** to 495  $\mu\text{L}$  **HRP-Conjugate Diluent**.
- 8.2.4 Mix thoroughly and gently. Hold no longer than 2 hours prior to using in procedure. Do not store at 1X concentration for future use.

## 8.3 **Microplate Preparation**

- Micro-plates are provided ready to use and do not require rinsing or blocking.
- Unused well strips should be returned to the original packaging, sealed and stored at 4°C.
- Equilibrate microplates to ambient temperatures prior to opening to reduce potential condensation.

## 8.4 **1X Wash Buffer**

- 8.4.1 If crystals have formed in the 20X **Wash Buffer** concentrate, equilibrate to room temperature and mix gently until crystals have completely dissolved.
- 8.4.2 Add the entire 15 mL contents of the 20X **Wash Buffer** bottle to 285 mL of ultra-pure water to a clean > 1,000 mL bottle or other vessel.
- 8.4.3 Seal and mix gently by inversion. Avoid foaming or bubbles.
- 8.4.4 Store the **1X Wash Buffer** at room temperature until ready to use in the procedure. Store the prepared **1X Wash Buffer** at 4°C for no longer than 1 week. Do not freeze.

## 9. Sample Preparation

### 9.1 Sample Preparation and Storage

- Store samples to be assayed at 2-8°C for 24 hours prior being assayed.
- For long term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.
- Samples not indicated in the manual must be tested to determine if the kit is valid.
- Prepare samples as follows:
  - **Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature or overnight at 4°C before centrifugation for 15 minutes at 1,000 x g. Remove serum and assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles.
  - **Plasma** - Collect plasma using EDTA, or heparin as an anticoagulant. Centrifuge for 15 minutes at 1,000 x g at 2-8°C within 30 minutes of collection. Assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles.
  - **Tissue homogenates** - The preparation of tissue homogenates will vary depending upon tissue type. For this assay, tissue was rinsed with 1X PBS to remove excess blood, homogenized in 20 mL of 1X PBS and stored overnight at  $\leq -20^{\circ}\text{C}$ . After two freeze-thaw cycles were performed to break the cell membranes, the homogenates were centrifuged for 5 minutes at 5,000 x g. Remove the supernatant and assay immediately or aliquot and store at  $\leq -20^{\circ}\text{C}$ .

### 9.2 Sample Dilution

Target protein concentration must be estimated and appropriate sample dilution selected such that the final target protein concentration falls near the middle of the assay linear dynamic range.

- Dilute samples using **Sample Diluent**.
- Mix diluted samples gently and thoroughly.
- Pipetting less than 2  $\mu\text{L}$  is not recommended for optimal assay accuracy.
- Optimal dilution must be determined by the user according to their specific samples.



## 10. Assay Procedure

- Equilibrate all reagents and materials to ambient room temperature prior to use in the procedure.
- For optimal control of small potential variations in micro well-plate and day to day ambient temperature fluctuations, equilibrate all reagents prior to use and perform all incubation steps at 37°C.

- 10.1** Determine the required number of wells and return any remaining unused wells and desiccant to the pouch.
- 10.2** Retain at least one well as an absolute Blank without any samples or reagents.
- 10.3** Add 50 µL of serially titrated standards, diluted samples or blank into wells of the **LEP Microplate**. At least two replicates of each standard, sample or blank is recommended.
- 10.4** Immediately add 50 µL of **1X HRP Conjugated LEP Detector Antibody** to each well (excluding absolute Blank).
- 10.5** Cover the plate with the well plate lid and incubate at 37°C for 60 minutes.
- 10.6** Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.
- 10.7** Gently blot any remaining liquid from the wells by tapping inverted on the bench top onto paper toweling. Do not allow the wells to completely dry at any time
- 10.8** Wash plate three times with **1X Wash Buffer** as follows:
  - 10.8.1 Add 300 µL of **1X Wash Buffer** to each assay well.
  - 10.8.2 Incubate for 10 seconds.
  - 10.8.3 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.
  - 10.8.4 Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.
  - 10.8.5 Repeat steps 10.8.1 through 10.8.4 **two** more times.
- 10.9** Add 50µl of Substrate A and 50µl of Substrate B to each well, mix well. Incubate for 15 minutes at 37°C. Keeping the plate away from drafts and other temperature fluctuations in the dark.
- 10.10** Add 50 µL of **Stop Solution** to each well. Well color should change to gradations of yellow immediately. Add the **Stop Solution** in the same well order as done for the **TMB Substrate**.
- 10.11** Read the O.D. absorbance at 450 nm with a standard microplate reader within 5 minutes of stopping the reaction in step 10.10. If wavelength correction is available, set to 540 nm or 570 nm.

## 11. Calculation of Results

For analysis of the assay results, calculate the **Relative OD<sub>450</sub>** for each test or standard well as follows:

$$(\text{Relative OD}_{450}) = (\text{Well OD}_{450}) - (\text{Mean Blank Well OD}_{450})$$

The standard curve is generated by plotting the mean replicate **Relative OD<sub>450</sub>** of each standard serial dilution point vs. The respective standard concentration. The LEP concentration contained in the samples can be interpolated by using linear regression of each mean sample **Relative OD<sub>450</sub>** against the standard curve. This is best achieved using curve fitting software.

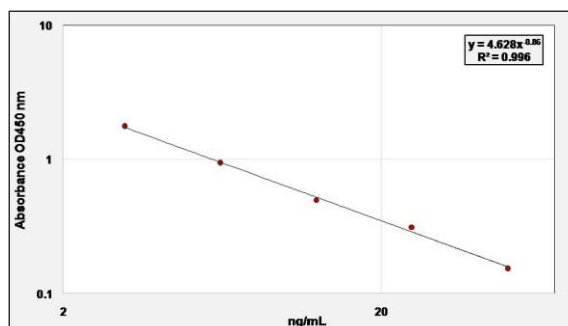
**Note:** if wavelength correction readings are available, subtract the readings at 540 nm or 570 nm from the readings at 450 nm. This may provide greater reading accuracy.

**Note:** if the samples measured were diluted, multiply the derived mean sample concentration by the dilution factor for a final sample concentration.

## 12. Typical Expected Data

### 12.1 Typical standard curve

This standard curve is for demonstration purposes only. An assay specific standard curve should be performed with each assay.



ng/mL	Absorbance (OD450 nm)		Mean
	1	2	
50	0.155	0.151	0.153
25	0.316	0.309	0.313
12.5	0.491	0.502	0.497
6.25	0.964	0.912	0.938
3.125	1.807	1.711	1.759

### 12.2 Reproducibility

Intra-assay reproducibility was evaluated with 20 replicates of 3 samples representing low, middle and high level LEP. Inter-assay reproducibility was evaluated with 3 samples representing low, middle and high level LEP using 8 replicates on each of 3 plates.

Intra-Assay CV <10%

Inter-Assay CV <12%

## 13. Technical Resources

### Technical Support:

For optimal service please be prepared to supply the lot number of the kit used.

#### USA

Aviva Systems Biology, Corp.  
5754 Pacific Center Blvd, Suite 201  
San Diego, CA 92121

Phone: 858-552-6979  
Toll Free: 888-880-0001  
Fax: 858-552-6975

Technical support: [techsupport@avivasysbio.com](mailto:techsupport@avivasysbio.com)

#### China

Beijing AVIVA Systems Biology  
6th Floor, B Building, Kaichi Tower  
#A-2 Jinfu Road.  
Daxing Industrial Development Zone  
Beijing, 102600, CHINA

Phone: (86)10-60214720  
Fax: (86)10-60214722  
E-mail: [support@avivasysbio.com.cn](mailto:support@avivasysbio.com.cn)

中国地址: 北京大兴工业开发区金辅路甲 2 号凯驰大厦 B 座 6 层 (102600)  
电话: 010-60214720/21  
传真: 010-60214722

产品售前咨询及销售: [sales@avivasysbio.com.cn](mailto:sales@avivasysbio.com.cn)  
售后及技术支持: [support@avivasysbio.com.cn](mailto:support@avivasysbio.com.cn)