For the quantitative measurement of Human Metallothionein in serum, plasma, tissue homogenates, cell culture supernatant, saliva and urine.

This product is intended for research use only.
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1. Background

**Principle**

Aviva Systems Biology Metallothionein ELISA Kit (Human) (OKCA00004) is based on standard sandwich enzyme-linked immuno-sorbent assay technology. An antibody specific for human Metallothionein has been pre-coated onto a 96-well plate (12 x 8 Well Strips). Standards or test samples are added to the wells, incubated and removed. A biotinylated detector antibody specific for human Metallothionein is added, incubated and followed by washing. Avidin-Biotin-Peroxidase Complex is then added, incubated and unbound conjugate is washed away. An enzymatic reaction is produced through the addition of TMB substrate which is catalyzed by HRP to generating a blue color product that changes to yellow after adding acidic stop solution. The density of yellow coloration read by absorbance at 450 nm and is quantitatively proportional to the amount of sample human Metallothionein captured in well.

2. Assay Summary
3. Precautions

• Read instructions fully prior to beginning use of the assay kit.
• Any deviations or modifications from the described method or use of other reagents could result in a reduction of performance.
• Reduce exposure to potentially harmful substances by wearing personal protective lab equipment including lab coats, gloves and glasses.
• For information on hazardous substances included in the kit please refer to the Material Safety Data Sheet (MSDS).
• Kit cannot be used beyond the expiration date on the label.

4. Storage and Stability

• Upon receipt store kit at 4°C for 6 months or -20°C for 12 months. Avoid multiple freeze/thaw cycles.

5. Kit Components

• The following reagents are the provided contents of the kit.

<table>
<thead>
<tr>
<th>Description</th>
<th>Quantity</th>
<th>Storage Conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-MT Microplate</td>
<td>96 Well (12 x 8 Well Strips)</td>
<td>2-8°C for 6 Months</td>
</tr>
<tr>
<td>MT Standard (Lyophilized)</td>
<td>2 x 400 pg</td>
<td>2-8°C for 6 Months</td>
</tr>
<tr>
<td>100X Biotinylated Anti-MT Detector Antibody</td>
<td>1 x 120 µL</td>
<td>-20°C long term</td>
</tr>
<tr>
<td>100X HRP-Avidin</td>
<td>1 x 120 µL</td>
<td>Avoid freeze-thaw</td>
</tr>
<tr>
<td>Detection Antibody Diluent</td>
<td>1 x 15 mL</td>
<td>2-8°C for 6 Months</td>
</tr>
<tr>
<td>HRP-Avidin Diluent</td>
<td>1 x 15 mL</td>
<td>2-8°C for 6 Months</td>
</tr>
<tr>
<td>Sample Diluent</td>
<td>1 x 50 mL</td>
<td></td>
</tr>
<tr>
<td>25X Wash Buffer</td>
<td>1 x 20 mL</td>
<td></td>
</tr>
<tr>
<td>TMB Substrate</td>
<td>1 x 10 mL</td>
<td></td>
</tr>
<tr>
<td>Stop Solution</td>
<td>1 x 10 mL</td>
<td></td>
</tr>
</tbody>
</table>

6. Required Materials Not Supplied

• Microplate reader capable of reading absorbance at 450 nm.
• Automated plate washer (optional).
• Pipettes capable of precisely dispensing 0.5 µL through 1 mL volumes of aqueous solutions.
• Pipettes or volumetric glassware capable of precisely measuring 1 mL through 100 mL of aqueous solutions.
• New, clean tubes and/or micro-centrifuge tubes for the preparation of standards or samples.
• Absorbent paper or paper toweling.
• Distilled or deionized ultrapure water.
• 37°C Incubator (optional).
7. Technical Application Tips

- Do not mix or substitute components from other kits.
- To ensure the validity of experimental operation, it is recommended that pilot experiments using standards and a small selection of sample dilutions to ensure optimal dilution range for quantitation.
- Samples exhibiting OD measurements higher than the highest standard should be diluted further in the appropriate sample dilution buffers.
- Prior to using the kit, briefly spin component tubes to collect all reagents at the bottom.
- Replicate wells are recommended for standards and samples.
- Cover microplate while incubating to prevent evaporation.
- Do not allow the microplate wells dry at any point during the assay procedure.
- Do not reuse tips or tube to prevent cross contamination.
- Avoid causing bubbles or foaming when pipetting, mixing or reconstituting.
- Completely remove of all liquids when washing to prevent cross contamination.
- Prepare reagents immediately prior to use and do not store, with the exception of the top standard.
- Equilibrate all materials to ambient room temperature prior to use (standards exception).
- For optimal results for inter- and intra-assay consistency, equilibrate all materials to 37°C prior to performing assay (standards exception) and perform all incubations at 37°C.
- Pipetting less than 1 µL is not recommended for optimal assay accuracy.
- Once the procedure has been started, all steps should be completed without interruption. Ensure that all reagents, materials and devices are ready at the appropriate time.
- Incubation times will affect results. All wells should be handled in the same sequential order and time intervals for optimal results.
- Samples containing precipitates or fibrin strands or which are hemolytic or lipemic might cause inaccurate results due to interfering factors.
- TMB Substrate is easily contaminated and should be colorless or light blue until added to plate. Handle carefully and protect from light.

8. Reagent Preparation

- Equilibrate all materials to room temperature prior to use and use prepare immediately prior to use.

8.1 Microplate Preparation

- Micro-plates are provided ready to use and do not require rinsing or blocking.
- Unused well strips should be returned to the original packaging, sealed and stored at 4°C.
- Equilibrate microplates to ambient temperatures prior to opening to reduce potential condensation.

8.2 1X Wash Buffer

8.2.1 If crystals have formed in the 25X Wash Buffer concentrate, equilibrate to room temperature and mix gently until crystals have completely dissolved.

8.2.2 Add the entire 20 mL contents of the 25X Wash Buffer bottle to 480 mL of ultra-pure water to a clean > 500 mL bottle or other vessel.

8.2.3 Seal and mix gently by inversion. Avoid foaming or bubbles.

8.2.4 Store the 1X Wash Buffer at room temperature until ready to use in the procedure. Store the prepared 1X Wash Buffer at 4°C for no longer than 1 week. Do not freeze.
8.3 Human Metallothionein Assay Standards

8.3.1 Prepare the human Metallothionein standards no greater than 2 hours prior to performing experiment. Standards should be held on ice until use in the experiment.

8.3.2 Reconstitute one vial of the provided 0.4 ng Lyophilized Metallothionein Standard for each experiment. Prepare the stock 400 pg/mL Metallothionein Standard by reconstituting one tube of Lyophilized Metallothionein Standard as follows:

8.3.2.1 Gently spin or tap the vial at 6,000 – 10,000 rpm for 30 seconds to collect all material at the bottom.

8.3.2.2 Add 1 mL of Sample Diluent to the vial.
8.3.2.3 Seal the vial then mix gently and thoroughly.
8.3.2.4 Leave the vial at ambient temperature for 15 minutes.

8.3.3 Prepare a set of seven serially diluted standards as follows:

8.3.3.1 Label tubes with numbers 2 – 8.

8.3.3.2 Use the undiluted 400 pg/mL Metallothionein Standard as the high standard point (Tube #1).

8.3.3.3 Add 300 µL of Sample Diluent to Tube #’s 2 – 8.

8.3.3.4 Prepare Standard #2 by adding 300 µL of 400 pg/mL Metallothionein (Tube #1) to Tube #2. Mix gently and thoroughly.

8.3.3.5 Prepare Standard #3 by adding 300 µL of Standard #2 from Tube #2 to Tube #3. Mix gently and thoroughly.

8.3.3.6 Prepare further serial dilutions through Tube #7. Reference the table below as a guide for serial dilution scheme.

8.3.3.7 Tube #8 is a blank standard (only Sample Diluent), which should be included with every experiment.

<table>
<thead>
<tr>
<th>Standard Number (Tube)</th>
<th>Standard To Dilute</th>
<th>Volume Standard to Dilute (µL)</th>
<th>Volume Sample Diluent Buffer (µL)</th>
<th>Total Volume (µL)</th>
<th>Final Concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>400 pg/mL Reconstituted Metallothionein Standard</td>
<td>1,000</td>
<td>NA</td>
<td>NA</td>
<td>400 pg/mL</td>
</tr>
<tr>
<td>2</td>
<td>400 pg/mL</td>
<td>300</td>
<td>300</td>
<td>600</td>
<td>200 pg/mL</td>
</tr>
<tr>
<td>3</td>
<td>200 pg/mL</td>
<td>300</td>
<td>300</td>
<td>600</td>
<td>100 pg/mL</td>
</tr>
<tr>
<td>4</td>
<td>100 pg/mL</td>
<td>300</td>
<td>300</td>
<td>600</td>
<td>50 pg/mL</td>
</tr>
<tr>
<td>5</td>
<td>50 pg/mL</td>
<td>300</td>
<td>300</td>
<td>600</td>
<td>25 pg/mL</td>
</tr>
<tr>
<td>6</td>
<td>25 pg/mL</td>
<td>300</td>
<td>300</td>
<td>600</td>
<td>12.5 pg/mL</td>
</tr>
<tr>
<td>7</td>
<td>12.5 pg/mL</td>
<td>300</td>
<td>300</td>
<td>600</td>
<td>6.25 pg/mL</td>
</tr>
<tr>
<td>8</td>
<td>NA</td>
<td>0</td>
<td>300</td>
<td>300</td>
<td>0.0 (Blank)</td>
</tr>
</tbody>
</table>
8.4 **1X Biotinylated Anti-Metallothionein Detector Antibody**

8.4.1 Prepare the **1X Biotinylated Anti-Metallothionein Detector Antibody** immediately prior to use by diluting the **100X Biotinylated Anti-Metallothionein Detector Antibody** 1:100 with **Antibody Diluent Buffer**.

8.4.2 For each well strip to be used in the experiment (8-wells) prepare 1,000 µL by adding 10 µL of **100X Biotinylated Anti-Metallothionein Detector Antibody** to 990 µL **Antibody Diluent Buffer**.

8.4.3 Mix thoroughly and gently. Hold no longer than 2 hours prior to using in procedure.

8.5 **1X HRP-Avidin**

8.5.1 Prepare the **1X HRP-Avidin** immediately prior to use by diluting the **100X HRP-Avidin** 1:100 with **HRP-Avidin Diluent**.

8.5.2 For each well strip to be used in the experiment (8-wells) prepare 1,000 µL by adding 10 µL of **100X HRP-Avidin** to 990 µL **HRP-Avidin Diluent**.

8.5.3 Mix thoroughly and gently. Hold no longer than 2 hours prior to using in procedure.
9. Sample Preparation

9.1 Sample Preparation and Storage

- Store samples to be assayed at 2-8°C for 24 hours prior to being assayed.
- For long term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.
- Samples not indicated in the manual must be tested to determine if the kit is valid.
- Prepare samples as follows:

  - Serum - Use a serum separator tube (SST) and allow samples to clot for two hours at room temperature or overnight at 4°C before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles.
  - Plasma - Collect plasma using EDTA, or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g, 2 - 8°C within 30 minutes of collection. Assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles. Centrifuge the sample again after thawing before the assay.
  - Cell Culture Supernatant - Remove particulates by centrifugation for 15 minutes at 1000 x g, 2 - 8°C and assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles.
  - Urine - Use a sterile container to collect urine samples. Remove any particulates by centrifugation for 15 minutes at 1,000 x g, 2 - 8°C and assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw cycles. Centrifuge again before assaying to remove any additional precipitates that may appear after storage.
  - Saliva - Remove particulates by centrifugation for 10 minutes at 4000 x g at 2 - 8°C and assay immediately or aliquot and store samples at -20°C. Avoid repeated freeze-thaw cycles. Recommend to use fresh saliva samples.

9.2 Sample Dilution

Target protein concentration must be estimated and appropriate sample dilution selected such that the final target protein concentration falls near the middle of the assay linear dynamic range.

- Recommended sample dilution for serum or plasma samples is 1:200 using Sample Diluent.
- The suggested 200-fold dilution can be prepared by adding 5 µL sample to 45 µL of Sample Diluent.
- Mix diluted samples gently and thoroughly.
- Pipetting less than 2 µL is not recommended for optimal assay accuracy.
10. Assay Procedure

• Equilibrate all reagents and materials to ambient room temperature prior to use in the procedure.
• Optionally, to control for small potential variations in micro well-plate and day to day ambient temperature fluctuations, equilibrate all reagents prior to use and perform all incubation steps at to 37°C for optimal consistency and reproducibility.

10.1 Determine the required number of wells and return any remaining unused wells and desiccant to the pouch.

10.2 Add 100 µL of serially titrated standards, diluted samples or blank into wells of the Anti-Metallothionein Microplate. At least two replicates of each standard, sample or blank is recommended.

10.3 Cover the plate with the well plate lid and incubate for 2 hours.

10.4 Remove the plate lid and discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.

10.5 Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.

10.6 Add 100 µL of prepared 1X Biotinylated Anti-Metallothionein Antibody to each well.

10.7 Cover with the well-plate lid and Incubate for 60 minutes.

10.8 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.

10.9 Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.

10.10 Wash plate 3 times with 1X Wash Buffer as follows:

10.10.1 Add 200 µL of 1X Wash Buffer to each assay well.
10.10.2 Incubate for 1 minute.
10.10.3 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle.
10.10.4 Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.
10.10.5 Repeat steps 10.10.1 through 10.10.4 two more times.

10.11 Add 100 µL of prepared 1X HRP-Avidin into each well and incubate for 60 minutes.

10.12 Discard the liquid in the wells by rigorously flicking into an acceptable waste receptacle or aspiration.

10.13 Gently blot any remaining liquid from the wells by tapping inverted on the benchtop onto paper toweling. Do not allow the wells to completely dry at any time.

10.14 Wash plate 5 times with 1X Wash Buffer as in Step 10.10.
10.15 Add 90 µL of **TMB Substrate** to each well and incubate in the dark for 15-30 minutes. Wells should change to gradations of blue. If the color is too deep, reduce the incubation time. 

*(NOTE: optimal incubation time must be determined by the user. Optimal development can be visualized by blue shading in the top four standard wells, while the remaining standards are still clear.)*

10.16 Add 50 µL of **Stop Solution** to each well. Well color should change to yellow immediately. Add the **Stop Solution** in the same well order as done for the **TMB Substrate**.

10.17 Read the O.D. absorbance at 450 nm with a standard microplate reader within 5 minutes of stopping the reaction in step 10.16. If wavelength correction is available, set to 540 nm or 570 nm.

### 11. Calculation of Results

For analysis of the assay results, calculate the **Relative OD** for each test or standard well as follows:

\[
(\text{Relative } \text{OD}_{450}) = (\text{Well } \text{OD}_{450}) - (\text{Mean Blank Well } \text{OD}_{450})
\]

The standard curve is generated by plotting the mean replicate **Relative OD** of each standard serial dilution point vs. the respective standard concentration. The human Metallothionein concentration contained in the samples can be interpolated by using linear regression of each mean sample **Relative OD** against the standard curve. This is best achieved using curve fitting software.

*Note:* if wavelength correction readings were available, subtract the readings at 540 nm or 570 nm from the readings at 450 nm. This may provide greater reading accuracy.

*Note:* if the samples measured were diluted, multiply the derived mean sample concentration by the dilution factor for a final sample concentration.

### 12. Typical Expected Data

#### 12.1 General Specifications

<table>
<thead>
<tr>
<th>General Specifications</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Range</strong></td>
<td>6.25 pg/ml-400 pg/ml</td>
</tr>
<tr>
<td><strong>LOD</strong></td>
<td>&lt; 0.813 pg/ml (Derived by linear regression of OD(_{450}) of the Mean Blank + 2xSD)</td>
</tr>
<tr>
<td><strong>Specificity</strong></td>
<td>Human Metallothionein</td>
</tr>
<tr>
<td></td>
<td>UniProt ID: P40731, Q86YXS,</td>
</tr>
<tr>
<td></td>
<td>GeneID: 4489</td>
</tr>
<tr>
<td><strong>Cross-Reactivity</strong></td>
<td>No detectable cross-reactivity with other relevant proteins</td>
</tr>
</tbody>
</table>

#### 12.2 Reproducibility

Three samples concentrations were measured in replicate within an assay plate and across replicate assays to assess Intra- and Inter-Assay precision.
12.3 **Typical absorbance values**

Expected absorbance for standards when TMB incubation is performed for 20 minutes at 37°C and measured at OD$_{450}$

<table>
<thead>
<tr>
<th>Sample Concentration (pg/mL)</th>
<th>Sample 1 Blank Subtracted (OD$_{450}$)</th>
<th>Sample 1 (OD$_{450}$)</th>
<th>Sample 2 (OD$_{450}$)</th>
<th>Mean (OD$_{450}$)</th>
<th>Sample 2 Blank Subtracted (OD$_{450}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blank</td>
<td>NA</td>
<td>0.114</td>
<td>0.118</td>
<td>0.116</td>
<td>0.209</td>
</tr>
<tr>
<td>6.25</td>
<td></td>
<td>0.321</td>
<td>0.329</td>
<td>0.325</td>
<td>0.569</td>
</tr>
<tr>
<td>12.5</td>
<td></td>
<td>0.415</td>
<td>0.407</td>
<td>0.411</td>
<td>0.295</td>
</tr>
<tr>
<td>25</td>
<td></td>
<td>0.681</td>
<td>0.689</td>
<td>0.685</td>
<td>0.569</td>
</tr>
<tr>
<td>50</td>
<td></td>
<td>1.079</td>
<td>1.071</td>
<td>1.075</td>
<td>0.959</td>
</tr>
<tr>
<td>100</td>
<td></td>
<td>1.488</td>
<td>1.486</td>
<td>1.487</td>
<td>1.371</td>
</tr>
<tr>
<td>200</td>
<td></td>
<td>1.867</td>
<td>1.857</td>
<td>1.862</td>
<td>1.746</td>
</tr>
<tr>
<td>400</td>
<td></td>
<td>2.110</td>
<td>2.104</td>
<td>2.107</td>
<td>1.991</td>
</tr>
</tbody>
</table>

12.4 **Typical standard curve**

This standard curve is for demonstration purposes only. An assay specific standard curve should be performed with each assay.

![Typical standard curve](image)

12.5 **Linearity**

Sample matrices (indicated below) were spiked with known concentrations of human Metallothionein, diluted to within the dynamic range of the assay and measured to assess the linearity of the assay measurements across the range of dilution points.

<table>
<thead>
<tr>
<th>Sample Type</th>
<th>Sample Dilution</th>
<th>Average Recovery</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human Serum</td>
<td>1:50</td>
<td>90 %</td>
<td>82-101%</td>
</tr>
<tr>
<td>(n=4)</td>
<td>1:100</td>
<td>94%</td>
<td>90-99%</td>
</tr>
<tr>
<td></td>
<td>1:200</td>
<td>93%</td>
<td>85-97%</td>
</tr>
</tbody>
</table>
13. Technical Resources

Technical Support:

For optimal service please be prepared to supply the lot number of the kit used.

USA

Aviva Systems Biology, Corp.
5754 Pacific Center Blvd, Suite 201
San Diego, CA 92121

Phone: 858-552-6979
Toll Free: 888-880-0001
Fax: 858-552-6975

Technical support: techsupport@avivasysbio.com

China

Beijing AVIVA Systems Biology
6th Floor, B Building, Kaichi Tower
#A-2 Jinfu Road.
Daxing Industrial Development Zone
Beijing, 102600, CHINA

Phone: (86)10-60214720
Fax: (86)10-60214722
E-mail: support@avivasysbio.com.cn

中国地址：北京大兴工业开发区金辅路甲2号凯驰大厦B座6层 (102600)
电话: 010-60214720/21
传真: 010-60214722

产品售前咨询及销售: sales@avivasysbio.com.cn
售后及技术支持: support@avivasysbio.com.cn